

## Effect of Petai Skin Extract Gel Administration on MITF Gene Expression and Melanin Count (in Vivo Experimental Study on Hyperpigmented Wistar Mice Exposed to UVB Exposed Hyperpigmentation)



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**ABSTRACT:** Prolonged exposure to high intensity UVB rays can induce the formation of Reactive Oxygen Species (ROS) which causes MITF activation thereby inducing melanin synthesis. Compounds contained in petai peel extract are known to play a role in inhibiting ROS production due to exposure to UVB rays. This study aims to determine the effect of administering petai peel extract gel on MITF gene expression and the amount of melanin in the skin tissue of mice exposed to high-intensity UVB. The research design was posttest only control group with a completely randomized design method. The samples studied were 24 mice exposed to UVB light with a wavelength of 302 nm and an energy of 390 mJ/cm<sup>2</sup>/day 3 times a week for 2 weeks. This research was carried out in four groups, namely the healthy group (K1), the negative control group (K2), treatment 1 (K3) with 10% petai peel extract gel and treatment 2 (K4) with 20% petai peel extract gel. MITF gene expression was analyzed using qRT-PCR and melanin was observed by specific staining with fontana-masson. qRT-PCR analysis showed that there was a significant decrease in MITF gene expression between groups K4 and K3 compared to group K2. Analysis of the amount of melanin also showed that there was a significant decrease in the mean in the K3 and K4 groups compared to K2 with a p value <0.05. Administration of petai peel extract gel can reduce the expression of the MITF gene and the amount of melanin in hyperpigmentation mice exposed to UVB light.

**KEYWORDS:** UVB exposure, petai peel extract, MITF, melanin

### I. INTRODUCTION

Ultraviolet B (UVB) radiation is the main factor causing hyperpigmentation, cessation of collagen synthesis and inflammation of the skin.<sup>1-3</sup> Hyperpigmentation is a condition in which there is an increase in the amount of melanin in the skin which can be visually observed in the presence of brownish pigment then blackening.<sup>4,5</sup> Hyperpigmentation caused by exposure to UVB light is controlled by certain genes, proteins, and enzymes, such as tyrosinase, a transcription factor associated with *melanocyte inducing transcription factor (MITF)*, tyrosinase-related protein 1 (TRP-1), and dopachrome tautomerase (TRP-2).<sup>6</sup> Activation of MITF leads to increased excessive melanin production in the skin.<sup>7</sup>

Increased exposure to UVB rays due to ozone layer damage in recent years and lifestyle changes have become important concerns as they can induce hyperpigmentation that increases cases of skin aging.<sup>1-3</sup> People with fair skin and older age have a higher risk of the effects of hyperpigmentation.<sup>4,5</sup> Although little research has been done on the specific incidence of hyperpigmentation, the incidence of melasma in men is 10%, while women prevalence is greater, especially at the age of 20-45 years. The ratio of melasma cases in women and men in Indonesia is 24:1.<sup>5</sup> Among the most common symptoms of aging, hyperpigmentation and facial wrinkles are the most common, this condition can affect a person's social interactions.<sup>6</sup> Until now, treatment with several chemical agents such as arbutin, azelaic acid, kojic acid, and hydroquinone was the main choice for preventing or treating skin hyperpigmentation and inducing collagen synthesis.<sup>7,8</sup> 9-11 9-11 12,13

Traditionally, petai seeds have been used by local people as a treatment for various ailments, especially skin diseases.<sup>14,15</sup> Based on the explanation above, petai seeds have been widely used. However, petai skin by the community is just thrown away and has

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not been widely used. Therefore, petai skin is used in the form of innovation into petai skin extract gel which acts as an antioxidant and anti-inflammatory. *Petai skin* also contains various types of polyphenols and flavonoids which are believed to be potential sources of bioactive compounds that are beneficial to health.<sup>16</sup> According to Isromi,<sup>12</sup> Petai bark (*Parkia speciosa*) contains flavonoids, phenols, and tannins that can be used as topical anti-inflammatories.<sup>13</sup> According to Rianti,<sup>17</sup> Petai skin contains flavonoids and phenols that can act as antioxidants. Other studies have also reported that petai has antioxidant properties<sup>18-20</sup> and anti-inflammatory.<sup>16,21</sup>

Previous studies have also reported that hyperpigmentation caused by exposure to UVB rays is regulated by various genes, proteins, and enzymes such as tyrosinase, and MITF.<sup>22</sup> In the melanin production pathway, tyrosinase plays a role in converting L-tyrosine to L-3,4-dihydroxyphenylalanine (L-DOPA).<sup>23</sup> L-DOPA is then oxidized to *L-DOPAquinone*, which then forms eumelanin and pheomelanin.<sup>24</sup> Other studies have also shown that chronic UVB exposure can cause oxidative stress, which activates phosphorylation of *mitogen-activated protein kinases* (MAPK) such as ERK, JNK, and p53 that accelerate melanin production.<sup>26,29</sup> Activation of these pathways increases transcription of MITF, which is a key factor in the activation of the enzymes tyrosinase, TRP-1, and TRP-2, which play a role in melanin production.<sup>29,30</sup> The flavonoid content in petai extract can inhibit ROS, which leads to suppression of MITF expression.<sup>31,32</sup> Other studies report that flavonoid compounds can increase the expression of antioxidants such as *superoxide dismutase* (SOD) so that it can inhibit the MITF pathway causing melanin production to decrease.<sup>33</sup> Flavonoid compounds contained in petai skin extract are thought to have the ability to reduce melanin production through antioxidant mechanisms.<sup>34,35</sup> However, studies on the effect of topical gels of petai skin extract on melanin and MITF in hyperpigmentation models due to UVB exposure have not been conducted. Based on this background, the researchers wanted to analyze the effect of petai skin extract in the form of a topical gel on MITF gene expression and the amount of melanin in the skin of hyperpigmented mice exposed to UVB.

## II. MATERIAL AND METHOD

### Study Design and Experimental Animals

The study was a post test only control group with a randomized design method complete with five repetitions per treatment. The study subjects used male rats of the wistar strain, aged 2-3 months with a body weight of 200-250 grams that fit the inclusion and inclusion criteria, adapted for 5 days. This study used 4 treatment groups, a group of healthy mice without UVB irradiation (K1), a negative control group, UVB-induced mice with base gel treatment (K2), a UVB-induced rat treatment group with a 10% topical dose of petai skin extract gel treatment (K3), and a UVB-induced rat treatment group with a 20% topical dose of petai skin extract gel treatment (K4).

### Research Materials

Research materials consist of materials for treatment such as water base gel, ketamine, xylazine, ethanol, aquades, rat feed, and chloroform.

### Research Equipment

This research uses several equipment to make animal models including UV light (broadband with a peak emission of 302 nm) with an energy of 390 mJ/cm<sup>2</sup>, razors, exposure cages, maintenance cages, drinking water for rats and hair cutters. The tools used for data collection are vacutainer, hematocrit tube, 5 mL pot, 6 mm biopsy punch, centrifuge, micropipette, 1000 uL micropipette tip, and 1.5 mL vial tube. Tools used for data analysis include microplate readers, microscopes, staining jars, coated desk glass, cover glass, and laptops.

### How to Make Petai Skin Extract Gel

±600 grams of petai peel is cut into small pieces, dried at 50-60°C and mashed into a dry powder. The process of making petai bark extract uses maceration techniques. Dried coolie petai powder is extracted using 70% ethanol then filtered and the filtrate is accommodated, the residue then macerated again by the same method. The choice of this solvent is because ethanol is able to filter active ingredients that are polar, semipolar and nonpolar. It is expected to produce an optimal amount of extract. The ethanol content is evaporated using a rotary evaporator to obtain a viscous extract. The results of evaporation are then thickened using a waterbath. The resulting viscous extract is used for the manufacture of gels. Gelling is done by mixing a hydrogel gel base (Katecho) of 200 mg with thick petai peel extract at P1 as much as 10% and P2 as much as 20%. Furthermore, to remove the aroma on pete added essential oil of lemon fruit. Stirring is carried out under aseptic conditions to form a homogeneous mixture of characteristics observed under a microscope.

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**Table 1. Petai bark extract gel formula.**

Composition	Dosage 10%	Dosage 20%
Hydrogel (Catecho) <a href="https://www.katecho.com/hydrogel/">https://www.katecho.com/hydrogel/</a>	180 mg	160 mg
The mapi leather eprint	20 mg	40 mg

### Dosage Assignment

Before the study is carried out, the dosage to be used for research is first determined. Previous research using extract gels containing flavonoids at a dose of 20% for topical use can reduce melanin levels<sup>91</sup>. The use of petai skin extract gel was carried out every day as much as 200 mg / rat so that the extra dose of petai peel used was 20 mg of extract in 180mg of gel per rat for a dose of 10% and 40 mg of extract in 160 mg of gel per rat for a dose of 20% (Table 1)<sup>92</sup>.

### UV-B Display

(1) Mice that had been adapted for 1 week were anesthetized with a mixture of ketamine (60 mg / kgbb) and xylazine (20 mg / kgbb). (2) The hair on the dorsal part of the rat shaved clean with a size of 5x5 cm. (3) Rats' backs were exposed to UV light (broadband with peak emission 302 nm) with a minimum dose of erythema 390 mJ/cm<sup>2</sup> 15 minutes six times in two weeks. (4) Rat Behaviour 1 (10% dose skin extract gel) and Behavior 2 (20% dose skin extract gel) were then treated topically using a mapi skin extract gel given once a day for 14 days after UV-B irradiation.

### Paraffin Block Making

(1) Dehydration: input pieces of tissue in alcohol stratified from 30%, 40%, 60%, 50%, 70%, 80%, 90%, 96% (stratified) to remove fluid from the tissue. Put the tissue into the alcohol-xylol solution for 1 hour then put the tissue in the pure xylol solution for 2 x 2 hours. (2) Paraffinization and Embedding: tissue input in liquid paraffin for 2 x 2 hours. Wait until the paraffin solidifies, cut the tissue in paraffin 4 microns thick with microtomes. The result of the tissue piece is attached to a glass object that has previously been smeared with polylysine as an adhesive. Put the tissue on the glass of the deparaffination object in the incubator and heated to a temperature of 56-58°C until the paraffin melts.

### Validation and analysis of post-UVB Melanin painting and treatment

(1) Melanin painting is done using the Masson Fontana painting method with the following stages: (1) The tissue slide is deparaffinized. (2) The slide is incubated in Working Silver Solution for 2 minutes, then rinse with water. (3) The slide is incubated in a 0.1% solution of Gold chloride 10 minutes, then rinsed with water. (4) The slide is incubated in a 5% solution of Hypo for 5 minutes then rinsed with water. (5) The slide is incubated in Nuclear-fast red solution for 5 minutes, then rinsed with water. (6) The slide is carried out by dehydration process, then install the desk glass.

Analysis of the amount of melanin was carried out using a digital analysis method based on where the slides were photographed using an Olympus CX41 microscope (Olympus, Japan) at a magnification of 400 times using the Optilab Pro camera (Miconos, Indonesia). Each preparation was photographed as many as 3 fields of view using JPEG format. The photos were edited using Adobe Photoshop CS3 software version 10.0.1 (Adobe Systems Inc., San Jose, CA, U.S.A) to select the epidermis layer using the *Polygonal Lasso tool*. The calculation of epidermal area in *pixels* was done using ImageJ software version 1.47t (National Institutes of Health, Bethesda, MD) using the *red channel* in the *RGB stack* by setting the *threshold* to near the maximum. An area of the epidermis is necessary to normalize the amount of melanin. The calculation of the amount of melanin in *pixels* was done using ImageJ software version 1.47t using the *red channel* by setting the *threshold*. The normalized amount of melanin is calculated according to the following formula for per field of view:<sup>92</sup>.

### Quantitative Analysis of MITF Gene Expression using RT-PCR

(1) RNA extraction and cDNA synthesis: RNA isolation of skin tissue was performed using TRIzol<sup>®</sup> reagent, (Invitrogen Life Technologies) and cDNA manufacturing using iScript cDNA Synthesis Kit (Bio-Rad iScript gDNA Clear cDNA synthesis Kit Catalog) using Reverse Transcriptase PCR (RT-PCR) thermal cycler C1000 (Bio-Rad). (2) MITF Gene Expression Determination: MITF was amplified using PCR-RFLP Technique, using PCR 2x PCR Master mix solution (iNtRON,<sup>®</sup> catalog number 25027) in a 0.2 mL vial tube with a total volume of 50 uL for 1 sample. PCR was performed using the thermal cycle of DNA: Applied Veriti Biosystems 96.

**Table 2. PCR Mix CXCL8 Components.**

Component	Kind
Primer	Forward MITF 5'-GCTGGAGACGGAAGCTCTGCT-3'
	Reverse MITF

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	5'-GAGTGGGAGGGAGAGTGAGG-3'
Reagent	Trizol Reagen
Rna Transcribed	High Capacity cDNA Reverse Transcription
cDNA	SYBR Green

## Network Sampling and Storage

Mice after 24 hours of the last treatment were turned off by cervical dislocation for tissue retrieval. Skin tissue is taken using a 6 mm punch biopsy in the induced part of the skin. Skin tissue samples are preserved in a later RNA solution to maintain RNA quality. Skin samples in RNA are stored at -20oC until the PCR analysis process is carried out.

## III. RESULT

### Petai Skin Extraction

Petai peel extract in this study was obtained by maceration method using ethanol solvent and produced an extract yield of 8.00%. The phytochemical screening results of petai bark extract show that petai skin is positive for phenol, phenolic, tannin, flavonoid, terpenoid, and saponin group compounds (Appendix. In this study, the total flavonoids and phenolics in petai bark extract were also determined using spectrophotometric methods. In 1 gram of petai bark extract contains flavonoids of 65.27mg ± 1.20 and phenolics of 44.70mg± 1.22. These results prove that most of the compounds contained in petai bark extract are flavonoids.

### Hyperpigmentation Model Validation

In this study using a hyperpigmentation model. Animal models induced hyperpigmentation with 302 nm UVB irradiation with an energy intensity of 390mJ/cm2 for three times a week for two weeks. Validation of hyperpigmentation is observed on day 14. In coloring *Fontana masson* shows that there is a significant increase in melanin production which is characterized by brown pigment in the epidermis (melanocyte cells). In the group given UVB irradiation (negative control) the amount of melanin increased by 46.5% (Figure 1).

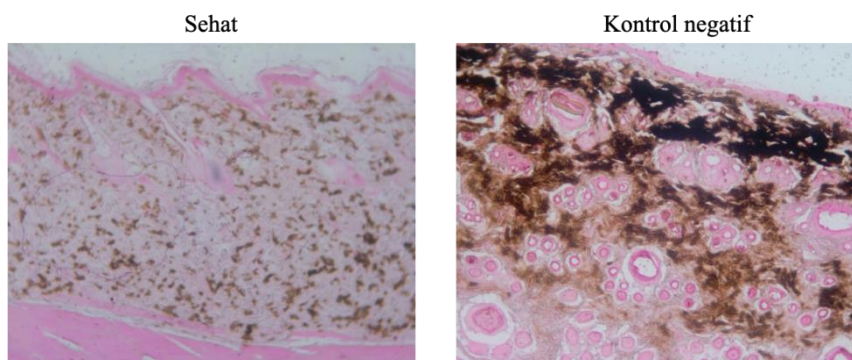


Figure 1. Validation of hyperpigmentation by masson fontane staining (A) Healthy mice and (B) UVB irradiated mice.

### Effects of 10% and 20% Doses of Petai Skin Extract Gel on MITF Gene Expression

In this study, researchers found that petai peel extract gel was able to reduce MITF gene expression and melanin count in hyperpygemntation model mice significantly dose-dependent (Table 3; Figure 2).

Table 3. Data from MITF and Melanin Gene Expression Research

Variable	Group			
	Sehat=5 Mean±SD K1	Negative control n=5 Mean±SD K2	Gel Extract Kulit Petai Dosage 10% n=5 Mean±SD K3	Gel Extract Kulit Petai Dosage 20% n=5 Mean±SD K4
Expressive has MITF	1.03±0.01	3.52±0.84	0.84±0.05	0.80±0.15
<i>Saphiro wilk</i>	0,415*	0,723*	0,181	0,362
<i>Levene test</i>				0,001**

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<i>One way ANOVA</i>					0,000***
Total Melanin	10.95±2.48	32.36±3.92	\$27.98±2.58	28.34±2.24	
<i>Saphiro wilk</i>	0,627	0,251	0,963	0,675	
<i>Levene test</i>				0,687	
<i>Kruskal-Wallis Test</i>					0,000***

Information:

\*Saphiro Wilk test ( $p < 0.05$  = abnormal)

\*\* Levene's Test ( $p < 0.05$  = not homogeneous)

Kruskal Wallis ( $p < 0.05$  = different meanings)

Based on the results of the study shown in table 3. The mean expression of the MITF gene in the gel group of petai skin extract dose was the lowest dose of 20% (K4) ( $0.80 \pm 0.15$ ), then followed by the average expression of the MITF gene in the gel group of petai skin extract dose of 10% (K3) ( $0.84 \pm 0.05$ ). The highest ratio in the negative control treatment group (K2) was  $3.52 \pm 0.84$ . The MITF gene expression data of all groups are normally distributed, shown by *Shapiro Wilk* results obtained a value of  $p > 0.05$  and also have inhomogeneous data variants indicated by the results of *Levene's Test* with a value of  $p = 0.001$  ( $p < 0.05$ ). The distribution and variant of MITF gene expression data are normal and inhomogeneous, so parametric statistical analysis with *One way-ANOVA* produces a value of  $p = 0.000$  ( $p < 0.05$ ) so that there is a significant difference in the average expression of MITF genes between the four groups. The significant *One way ANOVA* test results were followed by the *Tamhane Post Hoc* test to see which group was the most influential.

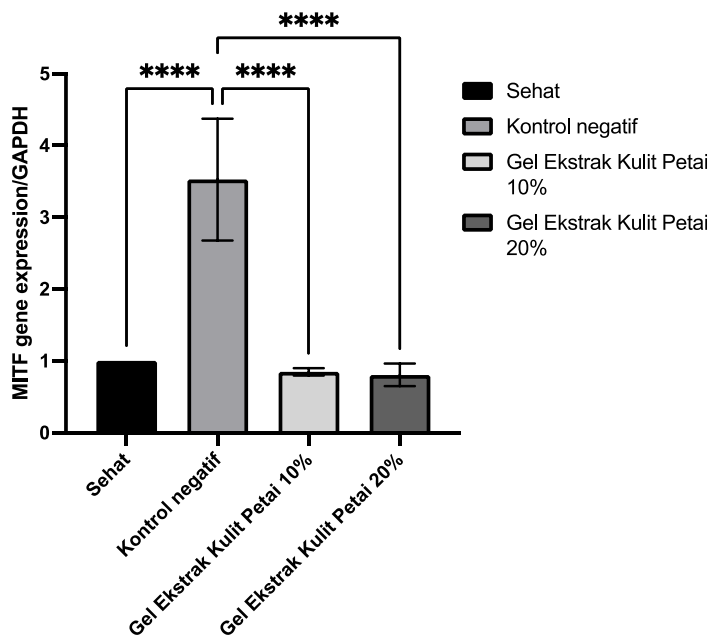


Figure 2. Graph of MITF gene expression across research groups

Table 4. Post Hoc Tamhane Test of MITF Gene Expression in Each Group

Group	Comparison Group	Say.	95% confidence interval	
			Lower Limit	Upper Limit
K1	K2	0,005*	-3,9532	-1,0502
	K3	0,002*	0,0891	0,2642
	K4	0,111	-0,0561	0,4883
K2	K3*	0,003*	1,2298	4,1269
	K4	0,003*	1,2936	4,1464
K3	K4*	0,993	-0,2182	0,3015

Sign \* ( $p < 0.05$ ) indicates a significantly different group

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Based on the data above, the average comparison between the K2 group with K3 (0.003) and K2 with K4 (0.003) showed a significant difference, while the comparison between the K3 and K4 groups (0.993) there was no significant difference. In the comparison of K1 and K2 obtained a value of 0.005 ( $p < 0.05$ ) so that there is a significant difference between the two groups. The most significant decrease in MITF gene expression was shown in the administration of 20% petai peel extract with a lower limit value of 1.2936 and an upper limit value of 4.1464. The results of the *post hoc tamhane* test on MITF gene expression data showed that the administration of petai skin extract gel that can reduce MITF gene expression in male rats wistar strain hyperpigmentation model.

### Effects of Petai Skin Extract Gel Doses of 10% and 20% on Melanin Expression

In this study, researchers found that petai skin extract gel was able to reduce the amount of melanin in hyperpigmentation mice significantly depending on dose (Table 3; Figure 3). Based on the results of the study shown in table 3. The average amount of melanin in the K3 group ( $27.98 \pm 2.58$ ) was the lowest, then followed by the average amount of melanin in the K4 group ( $28.34 \pm 2.24$ ). The highest amount of melanin data was in the negative control group of  $32.36 \pm 3.92$ . Data on the amount of melanin of all normally distributed groups ( $p > 0.05$ ) based on *Shapiro Wilk's results*. Based on homogeneity analysis with Levene test shows homogeneous data ( $p > 0.05$ ). The distribution of data on the amount of normal melanin and homogen so that parametric statistical analysis with *one way ANOVA* resulted in a value of  $p = 0.000$  ( $p < 0.05$ ) so that it was stated that there was a significant difference in the average amount of melanin between the four groups. The results of a *significant one-way ANOVA* test were followed by a *post hoc LSD* test to see which group was the most influential.

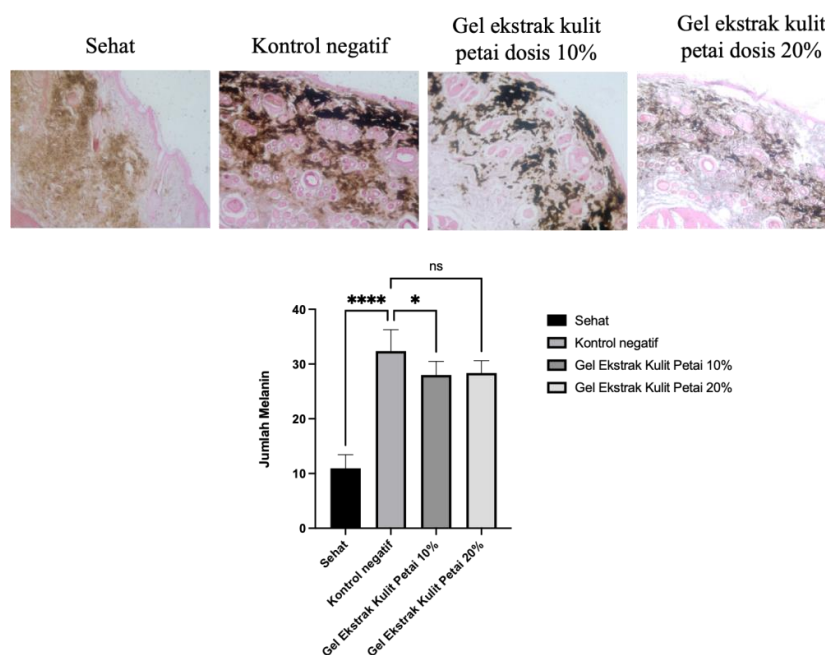


Figure 3. (A) Melanin staining picture in all treatment groups and (B) Graph of melanin count in all study groups

Table 5. *Post Hoc LSD* Test on Melanin Count in Each Group

Group	Comparison Group	Say.	95% confidence interval	
			Lower Limit	Upper Limit
K1	K2*	0,001*	-24,8654	17,9580
	K3	0,001*	-20,4904	-13,5830
	K4	0,001*	-20,8520	-13,9446
K2	K3*	0,016*	0,9213	7,8287
	K4*	0,025*	0,5596	7,4670
K3	K4	0,829	-3,8154	3,0920

Sign \* indicates a significantly different group.

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Based on the data above, an average comparison of K2 (negative control) with K3 (10% petai skin extract gel) (0.016) and K4 (20% petai skin extract gel) (0.025) was obtained, which means that there is a significant difference, while K3 and K4 (0.829) have no significant difference. In the comparison of groups K1 and K2 obtained a value of 0.001 ( $p < 0.05$ ) so that there is a significant difference between the two groups. The highest value of melanin amount was most significantly shown in the administration of 10% petai skin extract gel with a lower limit value of 0.9213 and an upper limit value of 7.8187. The results of the *LSD post hoc test* on melanin count data showed that the administration of petai skin extract gel that can reduce the amount of melanin in male rats of the Wistar strain hyperpigmentation model.

### IV. DISCUSSION

UVB exposure is a major risk factor for skin hyperpigmentation characterized by increased expression of melanin-forming proteins such as MITF.<sup>93</sup> UVB light irradiation induces DNA damage through the formation of oxidative ROS, resulting in activation of several melanogenesis pathways.<sup>94</sup> Recent research proves that petai bark extract which contains various secondary metabolite compounds such as flavonoids, alkaloids, tannins, and saponins can suppress ROS formation because it has antioxidant activity. The ability of extracts that can suppress ROS may prevent melanin production.<sup>35,95</sup> This study aimed to determine the effect of petai skin extract gel administration on MITF gene expression and melanin amount in hyperpigmented mouse models. This study used male rats of the wistar strain because it includes mammalian vertebrata with skin structures similar to human skin. Test animals were induced exposure to UVB light 302 nm with an energy intensity of 390mJ/cm<sup>2</sup> three times a week for 2 weeks<sup>93</sup>.

This study showed that MITF gene expression in the K3 (10% petai skin extract gel) and K4 (20% petai skin extract gel) groups decreased significantly compared to the hyperpigmented rat group that did not receive therapy (negative control). This suggests that decreased expression of the MITF gene can prevent hyperpigmentation. Flavonoid compounds derived from petai bark extract may inhibit TGF- $\beta$  thereby inhibiting MITF activity through inhibition of the PI3K/Akt pathway.<sup>96</sup> TGF- $\beta$  suppression has the potential to inhibit melanin biosynthesis in B16 melanoma cells. The ability of flavonoid compounds to inhibit TGF- $\beta$  has also been reported to inhibit the cAMP/protein kinase A pathway and induce GLI2, which then suppresses MITF, the central transcription factor of melanogenesis.<sup>97</sup> Meanwhile, through the SMAD pathway, TGF- $\beta$  suppression in inhibiting the process of melanogenesis by sending signals through ligand-specific heteromeric receptors, namely serine / threonine kinase receptors that function for phosphorylation and activation of (R)-Smad receptors, which leads to the formation of complexes with (Co)-Smad, Smad4, and transcriptional regulation of target genes so as to suppress MITF expression<sup>98</sup>.

This study further demonstrated the inhibitory effect of petai skin extract gel on the amount of melanin. This is due to petai bark extract which inhibits MITF so that it blocks TRP-1 activity and TRP-2 correlates with activation of eumelanin and pheomelanin formation pathways. Suppression of excessive TRP-1 expression can inhibit melanin synthesis<sup>99</sup>. Previous research reported that increased TRP-1 expression correlated with increased amounts of melanin due to UVB irradiation. However, increased TRP-2 expression is associated with melanoma cell proliferation<sup>100</sup>. In this study, the administration of petai skin extract gel significantly and dose-dependent decreased the amount of melanin. This shows that the administration of petai skin extract gel through MITF downregulation thus prevents melanin formation in melanocyte cells.

### V. CONCLUSION

1. Administration of petai bark extract at doses of 10% and 20% significantly decreased MITF gene expression in male rats of Wistar strain model UVB-induced hyperpigmentation.
2. Administration of petai bark extract at doses of 10% and 20% significantly decreased the amount of melanin in male rats of Wistar strain model of UVB-induced hyperpigmentation.

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### REFERENCES

- 1) Dale Wilson B, Moon S, Armstrong F. Comprehensive review of ultraviolet radiation and the current status on sunscreens. *J Clin Aesthet Dermatol*. 2012 Sep;5(9):18–23.
- 2) Merin KA, Shaji M, Kameswaran R. A Review on Sun Exposure and Skin Diseases. *Indian J Dermatol*. 2022;67(5):625.
- 3) Amaro-Ortiz A, Yan B, D’Orazio J. Ultraviolet Radiation, Aging and the Skin: Prevention of Damage by Topical cAMP Manipulation. *Molecules*. 2014 May 15;19(5):6202–19.

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- 4) Durai PC, Thappa DM, Kumari R, Malathi M. Aging in elderly: chronological versus photoaging. *Indian J Dermatol*. 2012 Sep;57(5):343–52.
- 5) Hughes MCB, Williams GM, Pigeon H, Fourtanier A, Green AC. Dietary Antioxidant Capacity and Skin Photoaging: A 15-Year Longitudinal Study. *Journal of Investigative Dermatology*. 2021 Apr;141(4):1111-1118.e2.
- 6) Samson N, Fink B, Matts PJ. Visible skin condition and perception of human facial appearance. *Int J Cosmet Sci*. 2010 Nov 3;32(3):167–84.
- 7) Solano F. Photoprotection and Skin Pigmentation: Melanin-Related Molecules and Some Other New Agents Obtained from Natural Sources. *Molecules*. 2020 Mar 27;25(7):1537.
- 8) Markiewicz E, Idowu O. <p>Melanogenic Difference Consideration in Ethnic Skin Type: A Balance Approach Between Skin Brightening Applications and Beneficial Sun Exposure</p>. *Clin Cosmet Investig Dermatol*. 2020 Mar; Volume 13:215–32.
- 9) Tangau MJ, Chong YK, Yeong KY. Advances in cosmeceutical nanotechnology for hyperpigmentation treatment. Vol. 24, *Journal of Nanoparticle Research*. Springer Science and Business Media B.V.; 2022.
- 10) Couteau C, Coiffard L. Overview of Skin Whitening Agents: Drugs and Cosmetic Products. *Cosmetics*. 2016 Jul 25;3(3):27.
- 11) Saeedi M, Khezri K, Seyed Zakaryaei A, Mohammadamini H. A comprehensive review of the therapeutic potential of  $\alpha$ -arbutin. *Phytotherapy Research*. 2021 Aug 16;35(8):4136–54.
- 12) Isromi T, Winahyu DA, Tutik T. Uji Efektivitas Ekstrak Kulit Petai (*Parkia Speciosa*) Sebagai Antiinflamasi Terhadap Tikus Putih (*Rattus Novergicus*) Jantan Galur Wistar Yang Di Induksi Karagenan. *Jurnal Ilmu Kedokteran dan Kesehatan*. 2023 Apr 9;10(3):1605–14.
- 13) Maulana I, Kurniati Roddu A, Suriani S. Uji Efektifitas Ekstrak Kulit Petai (*Parkia speciosa* Hassk) Terhadap Mencit (*Mus musculus*) Sebagai Anti Inflamasi. *Lambung Farmasi: Jurnal Ilmu Kefarmasian*. 2020 Jul 20;1(2):80.
- 14) Azemi AK, Nordin ML, Hambali KA, Noraldin NA, Mokhtar SS, Rasool AHG. Phytochemical Contents and Pharmacological Potential of *Parkia speciosa* Hassk. for Diabetic Vasculopathy: A Review. *Antioxidants*. 2022 Feb 21;11(2):431.
- 15) Kamisah Y, Othman F, Qodriyah HMS, Jaarin K. *Parkia speciosa* Hassk.: A Potential Phytomedicine. *Evidence-Based Complementary and Alternative Medicine*. 2013;2013:1–9.
- 16) Azemi AK, Nordin ML, Hambali KA, Noraldin NA, Mokhtar SS, Rasool AHG. Phytochemical Contents and Pharmacological Potential of *Parkia speciosa* Hassk. for Diabetic Vasculopathy: A Review. *Antioxidants (Basel)*. 2022 Feb 21;11(2).
- 17) Rianti A, Parassih EK, Novenia AE, Christpoher A, Lestari D, Kiyat W El. Potensi Ekstrak Kulit Petai (*Parkia speciosa*) Sebagai Sumber Antioksidan. *Jurnal Dunia Gizi*. 2018 Nov 15;1(1):10.
- 18) Saleh MSM, Jalil J, Zainalabidin S, Asmadi AY, Mustafa NH, Kamisah Y. Genus *Parkia*: Phytochemical, Medicinal Uses, and Pharmacological Properties. *Int J Mol Sci*. 2021 Jan 9;22(2):618.
- 19) Mustafa Khalid N, Babji AS. Antioxidative and Antihypertensive Activities of Selected Malaysian ulam (salad), Vegetables and Herbs. *J Food Res*. 2018 Mar 30;7(3):27.
- 20) Ghasemzadeh A, Jaafar HZE, Bukhori MFM, Rahmat MH, Rahmat A. Assessment and comparison of phytochemical constituents and biological activities of bitter bean (*Parkia speciosa* Hassk.) collected from different locations in Malaysia. *Chem Cent J*. 2018 Dec 7;12(1):12.
- 21) Gui JS, Jalil J, Jubri Z, Kamisah Y. *Parkia speciosa* empty pod extract exerts anti-inflammatory properties by modulating NF $\kappa$ B and MAPK pathways in cardiomyocytes exposed to tumor necrosis factor- $\alpha$ . *Cytotechnology*. 2019 Feb 1;71(1):79–89.
- 22) Wawrzyk-Bochenek I, Rahnema M, Stachura M, Wilczyński S, Wawrzyk A. Evaluation of the Reduction of Skin Hyperpigmentation Changes under the Influence of a Preparation Containing Kojic Acid Using Hyperspectral Imaging—Preliminary Study. *J Clin Med*. 2023 Apr 1;12(7).
- 23) Hori I, Nihei K ichi, Kubo I. Structural criteria for depigmenting mechanism of arbutin. *Phytotherapy Research*. 2004 Jun;18(6):475–9.
- 24) Galván I, Wakamatsu K, Alonso-Alvarez C, Solano F. Buthionine sulfoximine diverts the melanogenesis pathway toward the production of more soluble and degradable pigments. *Bioorg Med Chem Lett*. 2014 May;24(9):2150–4.
- 25) Cano M, Guerrero-Castilla A, Nabavi SM, Ayala A, Argüelles S. Targeting pro-senescence mitogen activated protein kinase (Mapk) enzymes with bioactive natural compounds. *Food and Chemical Toxicology*. 2019 Sep;131:110544.
- 26) Lee YI, Choi S, Roh WS, Lee JH, Kim TG. Cellular Senescence and Inflammation in the Skin Microenvironment. *Int J Mol Sci*. 2021 Apr 8;22(8):3849.



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- 27) López-Camarillo C, Aréchaga Ocampo E, López Casamichana M, Pérez-Plasencia C, Álvarez-Sánchez E, Marchat LA. Protein Kinases and Transcription Factors Activation in Response to UV-Radiation of Skin: Implications for Carcinogenesis. *Int J Mol Sci.* 2011 Dec 23;13(1):142–72.
- 28) Luchetti F, Betti M, Canonico B, Arcangeletti M, Ferri P, Galli F, et al. ERK MAPK activation mediates the antiapoptotic signaling of melatonin in UVB-stressed U937 cells. *Free Radic Biol Med.* 2009 Feb;46(3):339–51.
- 29) Lee SJ. Suppression of  $\alpha$ -MSH and IBMX-induced melanogenesis by cordycepin via inhibition of CREB and MITF, and activation of PI3K/Akt and ERK-dependent mechanisms. *Int J Mol Med.* 2011 Oct 3;
- 30) Kim DS, Park SH, Park KC. Transforming growth factor- $\beta$ 1 decreases melanin synthesis via delayed extracellular signal-regulated kinase activation. *Int J Biochem Cell Biol.* 2004 Aug;36(8):1482–91.
- 31) Hwang YP, Oh KN, Yun HJ, Jeong HG. The flavonoids apigenin and luteolin suppress ultraviolet A-induced matrix metalloproteinase-1 expression via MAPKs and AP-1-dependent signaling in HaCaT cells. *J Dermatol Sci.* 2011 Jan;61(1):23–31.
- 32) Butarbutar RH, Robiyanto R, Untari EK. Potensi Ekstrak Etanol Daun Petai (*Parkia speciosa* Hassk.) Terhadap Kadar Superoksida Dismutase (SOD) Pada Plasma Tikus yang Mengalami Stres Oksidatif. *Pharmaceutical Sciences and Research.* 2016 Aug;3(2):97–106.
- 33) Zhan JYX, Wang XF, Liu YH, Zhang ZB, Wang L, Chen JN, et al. Andrographolide sodium bisulfate prevents uv-induced skin photoaging through inhibiting oxidative stress and inflammation. *Mediators Inflamm.* 2016;2016.
- 34) Siow HL, Gan CY. Extraction of antioxidative and antihypertensive bioactive peptides from *Parkia speciosa* seeds. *Food Chem.* 2013;141(4):3435–42.
- 35) Izzah Ahmad N, Abdul Rahman S, Leong YH, Azizul NH. A Review on the Phytochemicals of *Parkia Speciosa*, Stinky Beans as Potential Phytomedicine. *J Food Sci Nutr Res.* 2019;02(03).
- 36) Iqbal IY. Pemberian krim ekstrak etanol biji petai (*parkia speciosa*) 20% sama efektif dengan krim Hidrokuinon 4% dalam menghambat Pembentukan jumlah melanin pada kulit Marmut (*cavia porcellus*) yang dipapar sinar Ultraviolet B. [Denpasar]: Universitas Udaya; 2019.
- 37) Wijayanti A. Karakteristik Ekstrak Kulit Petai (*Parkia speciosa* Hassk) dengan Pelarut Ethanol 70% dan etil Asetat. *Jurnal Ilmu Kesehatan Bhakti Setya Medika.* 2021 Dec 25;6(2):123–7.
- 38) Al Batran R, Al-Bayat F, Jamil Al-Obaidi MM, Abdulkader AM, Hadi HA, Ali HM, et al. In Vivo Antioxidant and Antiulcer Activity of *Parkia speciosa* Ethanolic Leaf Extract against Ethanol-Induced Gastric Ulcer in Rats. *PLoS One.* 2013 May 28;8(5):e64751.
- 39) Chen YS, Lee SM, Lin CC, Liu CY. Hispolon Decreases Melanin Production and Induces Apoptosis in Melanoma Cells through the Downregulation of Tyrosinase and Microphthalmia-Associated Transcription Factor (MITF) Expressions and the Activation of Caspase-3, -8 and -9. *Int J Mol Sci.* 2014 Jan 17;15(1):1201–15.
- 40) Wei B, Zhang YP, Yan HZ, Xu Y, Du TM. Cilostazol promotes production of melanin by activating the microphthalmia-associated transcription factor (MITF). *Biochem Biophys Res Commun.* 2014 Jan;443(2):617–21.
- 41) Hsiao JJ, Fisher DE. The roles of microphthalmia-associated transcription factor and pigmentation in melanoma. *Arch Biochem Biophys.* 2014 Dec;563:28–34.
- 42) Lee SE, Park SH, Oh SW, Yoo JA, Kwon K, Park SJ, et al. Beauvericin inhibits melanogenesis by regulating cAMP/PKA/CREB and LXR- $\alpha$ /p38 MAPK-mediated pathways. *Sci Rep.* 2018 Oct 8;8(1):14958.
- 43) D’Mello S, Finlay G, Baguley B, Askarian-Amiri M. Signaling Pathways in Melanogenesis. *Int J Mol Sci.* 2016 Jul 15;17(7):1144.
- 44) Baxter LL, Pavan WJ. The etiology and molecular genetics of human pigmentation disorders. *Wiley Interdiscip Rev Dev Biol.* 2013 May;2(3):379–92.
- 45) Videira IF dos S, Moura DFL, Magina S. Mechanisms regulating melanogenesis\*. *An Bras Dermatol.* 2013 Feb;88(1):76–83.
- 46) Hida T, Kamiya T, Kawakami A, Ogino J, Sohma H, Uhara H, et al. Elucidation of Melanogenesis Cascade for Identifying Pathophysiology and Therapeutic Approach of Pigmentary Disorders and Melanoma. *Int J Mol Sci.* 2020 Aug 25;21(17):6129.
- 47) Lee SJ, Lee WJ, Chang SE, Lee GY. Antimelanogenic effect of ginsenoside Rg3 through extracellular signal-regulated kinase-mediated inhibition of microphthalmia-associated transcription factor. *J Ginseng Res.* 2015 Jul;39(3):238–42.
- 48) Wang JH, Hwang SJ, Lee SK, Choi Y, Byun CK, Son CG. Anti-Melanogenic Effects of Fractioned *Cynanchum atratum* by Regulation of cAMP/MITF Pathway in a UVB-Stimulated Mice Model. *Cells.* 2023 May 14;12(10):1390.

## Effect of Petai Skin Extract Gel Administration on MITF Gene Expression and Melanin Count (in Vivo Experimental Study on Hyperpigmented Wistar Mice Exposed to UVB Exposed Hyperpigmentation)

- 49) Jablonski NG, Chaplin G. Human skin pigmentation as an adaptation to UV radiation. *Proceedings of the National Academy of Sciences*. 2010 May 11;107(supplement\_2):8962–8.
- 50) Lin JY, Fisher DE. Melanocyte biology and skin pigmentation. *Nature*. 2007 Feb 21;445(7130):843–50.
- 51) Kumari S, Thng S, Verma N, Gautam H. Melanogenesis Inhibitors. *Acta Dermato Venereologica*. 2018;98(10):924–31.
- 52) Pillaiyar T, Namasivayam V, Manickam M, Jung SH. Inhibitors of Melanogenesis: An Updated Review. *J Med Chem*. 2018 Sep 13;61(17):7395–418.
- 53) Nguyen NT, Fisher DE. MITF and UV responses in skin: From pigmentation to addiction. *Pigment Cell Melanoma Res*. 2019 Mar 3;32(2):224–36.
- 54) Vance KW, Goding CR. The Transcription Network Regulating Melanocyte Development and Melanoma. *Pigment Cell Res*. 2004 Aug;17(4):318–25.
- 55) Xu W, Gong L, Haddad MM, Bischof O, Campisi J, Yeh ETH, et al. Regulation of Microphthalmia-Associated Transcription Factor MITF Protein Levels by Association with the Ubiquitin-Conjugating Enzyme hUBC9. *Exp Cell Res*. 2000 Mar;255(2):135–43.
- 56) Alam MB, Bajpai VK, Lee J, Zhao P, Byeon JH, Ra JS, et al. Inhibition of melanogenesis by jineol from *Scolopendra subspinipes mutilans* via MAP-Kinase mediated MITF downregulation and the proteasomal degradation of tyrosinase. *Sci Rep*. 2017 Apr 10;7(1):45858.
- 57) Hong Y, Song B, Chen HD, Gao XH. Melanocytes and Skin Immunity. *Journal of Investigative Dermatology Symposium Proceedings*. 2015 Jul;17(1):37–9.
- 58) Nishio T, Usami M, Awaji M, Shinohara S, Sato K. Dual effects of acetylsalicylic acid on ERK signaling and Mitf transcription lead to inhibition of melanogenesis. *Mol Cell Biochem*. 2016 Jan 23;412(1–2):101–10.
- 59) Wellbrock C, Arozarena I. Microphthalmia-associated transcription factor in melanoma development and MAP-kinase pathway targeted therapy. *Pigment Cell Melanoma Res*. 2015 Jul 17;28(4):390–406.
- 60) Kawai T, Akira S. Signaling to NF- $\kappa$ B by Toll-like receptors. *Trends Mol Med*. 2007 Nov;13(11):460–9.
- 61) Thawabteh AM, Jibreen A, Karaman D, Thawabteh A, Karaman R. Skin Pigmentation Types, Causes and Treatment—A Review. *Molecules*. 2023 Jun 18;28(12):4839.
- 62) Moolla S, Miller-Monthrope Y. Dermatology: how to manage facial hyperpigmentation in skin of colour. *Drugs Context*. 2022 May 31;11:1–14.
- 63) Nguyen NT, Fisher DE. MITF and UV responses in skin: From pigmentation to addiction. *Pigment Cell Melanoma Res*. 2019 Mar;32(2):224–36.
- 64) Integrated Taxonomic Information System. Taxonomic Hierarchy : *Parkia speciosa* Hassk. 2023.
- 65) Choy KW, Murugan D, Leong XF, Abas R, Alias A, Mustafa MR. Flavonoids as Natural Anti-Inflammatory Agents Targeting Nuclear Factor-Kappa B (NF $\kappa$ B) Signaling in Cardiovascular Diseases: A Mini Review. *Front Pharmacol*. 2019 Oct 31;10.
- 66) Qin Z, Balimunkwe RM, Quan T. Age-related reduction of dermal fibroblast size upregulates multiple matrix metalloproteinases as observed in aged human skin *in vivo*. *British Journal of Dermatology*. 2017 Nov;177(5):1337–48.
- 67) Martinez-Esparza M, Jimenez-Cervantes C, Solano F, Lozano JA, Garcia-Borron JC. Mechanisms of melanogenesis inhibition by tumor necrosis factor- $\alpha$  in B16/F10 mouse melanoma cells. *Eur J Biochem*. 1998 Jul 1;255(1):139–46.
- 68) Kim DS, Park SH, Park KC. Transforming growth factor- $\beta$ 1 decreases melanin synthesis via delayed extracellular signal-regulated kinase activation. *Int J Biochem Cell Biol*. 2004 Aug;36(8):1482–91.
- 69) Liu F, Fu Y, Meyskens FL. MITF regulates cellular response to reactive oxygen species through transcriptional regulation of APE-1/Ref-1. *Journal of Investigative Dermatology*. 2009 Feb;129(2):422–31.
- 70) Kaminski K, Kazmierczak U, Kolenda T. Oxidative stress in melanogenesis and melanoma development. Vol. 26, *Wspolczesna Onkologia*. Termedia Publishing House Ltd.; 2022. p. 1–7.
- 71) Ishikawa Y, Bächinger HP. A molecular ensemble in the rER for procollagen maturation. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*. 2013 Nov;1833(11):2479–91.
- 72) Zhao W, Wang X, Sun KH, Zhou L.  $\alpha$ -smooth muscle actin is not a marker of fibrogenic cell activity in skeletal muscle fibrosis. *PLoS One*. 2018 Jan 10;13(1):e0191031.
- 73) Wölfle U, Esser PR, Simon-Haarhaus B, Martin SF, Lademann J, Schempp CM. UVB-induced DNA damage, generation of reactive oxygen species, and inflammation are effectively attenuated by the flavonoid luteolin *in vitro* and *in vivo*. *Free Radic Biol Med*. 2011;50(9):1081–93.

## Effect of Petai Skin Extract Gel Administration on MITF Gene Expression and Melanin Count (in Vivo Experimental Study on Hyperpigmented Wistar Mice Exposed to UVB Exposed Hyperpigmentation)

- 74) Arab Sadeghabadi Z, Abbasalipourkabir R, Mohseni R, Ziamajidi N. Investigation of oxidative stress markers and antioxidant enzymes activity in newly diagnosed type 2 diabetes patients and healthy subjects, association with IL-6 level. *J Diabetes Metab Disord*. 2019;18(2):437–43.
- 75) Ahmed SMU, Luo L, Namani A, Wang XJ, Tang X. Nrf2 signaling pathway: Pivotal roles in inflammation. *Biochim Biophys Acta Mol Basis Dis* [Internet]. 2017;1863(2):585–97. Available from: <http://dx.doi.org/10.1016/j.bbadis.2016.11.005>
- 76) Shin JM, Kim MY, Sohn KC, Jung SY, Lee HE, Lim JW, et al. Nrf2 negatively regulates melanogenesis by modulating PI3K/Akt signaling. *PLoS One*. 2014 Apr 24;9(4).
- 77) Garufi A, Pistritto G, D'orazi V, Cirone M, D'orazi G. The Impact of NRF2 Inhibition on Drug-Induced Colon Cancer Cell Death and p53 Activity: A Pilot Study. *Biomolecules*. 2022 Mar 1;12(3).
- 78) Addor FAS. Antioxidants in dermatology. *An Bras Dermatol*. 2017;92(3):356–62.
- 79) Arauz J, Ramos-Tovar E, Muriel P. Redox state and methods to evaluate oxidative stress in liver damage: From bench to bedside. *Ann Hepatol*. 2016;15(2):160–73.
- 80) Addor FAS. Antioxidants in dermatology. *An Bras Dermatol*. 2017;92(3):356–62.
- 81) A. Satyanarayana D, K. Kulkarni P, G. Shivakumar H. Gels and Jellies as a Dosage Form for Dysphagia Patients: A Review. *Curr Drug Ther*. 2011;6(2):79–86.
- 82) Bowlby M, Blume P, Schmidt B, Donegan R. Safety and efficacy of Becaplermin gel in the treatment of diabetic foot ulcers. *Chronic Wound Care Management and Research*. 2014;11.
- 83) Mayori H, . K, . M, Purnama D, Maulina Sari R. Systematic Review Efektivitas Limbah Kulit Petai (*Parkia speciosa* Hassk) sebagai Fitomedicine untuk Mengobati Masalah Kesehatan Tertentu. *JURNAL BIOSHELL*. 2023 Apr 27;12(1):66–76.
- 84) Varela MT, Ferrarini M, Mercaldi VG, Sufi B da S, Padovani G, Nazato LIS, et al. Coumaric acid derivatives as tyrosinase inhibitors: Efficacy studies through in silico, in vitro and ex vivo approaches. *Bioorg Chem*. 2020 Oct;103:104108.
- 85) Ke Y, Wang XJ. TGF $\beta$  Signaling in Photoaging and UV-Induced Skin Cancer. *Journal of Investigative Dermatology*. 2021 Apr;141(4):1104–10.
- 86) Pandel R, Poljšak B, Godic A, Dahmane R. Skin photoaging and the role of antioxidants in its prevention. *ISRN Dermatol*. 2013 Sep 12;2013:930164.
- 87) Speeckaert R, Van Gele M, Speeckaert MM, Lambert J, van Geel N. The biology of hyperpigmentation syndromes. *Pigment Cell Melanoma Res*. 2014 Jul;27(4):512–24.
- 88) Rinnerthaler M, Bischof J, Streubel M, Trost A, Richter K. Oxidative Stress in Aging Human Skin. *Biomolecules*. 2015 Apr 21;5(2):545–89.
- 89) Domaszewska-Szostek A, Puzianowska-Kuźnicka M, Kuryłowicz A. Flavonoids in Skin Senescence Prevention and Treatment. *Int J Mol Sci*. 2021 Jun 25;22(13):6814.
- 90) Wu PY, Lyu JL, Liu YJ, Chien TY, Hsu HC, Wen KC, et al. Fisetin regulates Nrf2 expression and the inflammation-related signaling pathway to prevent UVB-induced skin damage in hairless mice. *Int J Mol Sci*. 2017;18(10).
- 91) Zhu W, Gao J. The use of botanical extracts as topical skin-lightening agents for the improvement of skin pigmentation disorders. In: *Journal of Investigative Dermatology Symposium Proceedings*. Nature Publishing Group; 2008. p. 20–4.
- 92) Iqbal IY. Pemberian Krim Ekstrak Etanol Biji Petai (*Parkia Speciosa*) 20% Sama Efektif Dengan Krim Hidrokuinon 4% Dalam Menghambat Pembentukan Jumlah Melanin Pada Kulit Marmut (*Cavia Porcellus*) Yang Dipapar Sinar Ultraviolet B Irah Yunita Iqbal. Denpasar, Indonesia;
- 93) You YJ, Wu PY, Liu YJ, Hou CW, Wu CS, Wen KC, et al. Sesamol inhibited ultraviolet radiation-induced hyperpigmentation and damage in C57BL/6 mouse skin. *Antioxidants*. 2019;8(7):1–16.
- 94) Kim HY, Sah SK, Choi SS, Kim TY. Inhibitory effects of extracellular superoxide dismutase on ultraviolet B-induced melanogenesis in murine skin and melanocytes. *Life Sci* [Internet]. 2018;210:201–8. Available from: <https://doi.org/10.1016/j.lfs.2018.08.056>
- 95) Chhikara N, Devi HR, Jaglan S, Sharma P, Gupta P, Panghal A. Bioactive compounds, food applications and health benefits of *Parkia speciosa* (stinky beans): A review. Vol. 7, *Agriculture and Food Security*. BioMed Central Ltd.; 2018.
- 96) Gui JS, Jalil J, Jubri Z, Kamisah Y. *Parkia speciosa* empty pod extract exerts anti-inflammatory properties by modulating NF $\kappa$ B and MAPK pathways in cardiomyocytes exposed to tumor necrosis factor- $\alpha$ . *Cytotechnology*. 2019 Feb 15;71(1):79–89.
- 97) Lee SE, Park SH, Oh SW, Yoo JA, Kwon K, Park SJ, et al. Beauvericin inhibits melanogenesis by regulating cAMP/PKA/CREB and LXR- $\alpha$ /p38 MAPK-mediated pathways. *Sci Rep*. 2018;8(1):1–12.

## Effect of Petai Skin Extract Gel Administration on MITF Gene Expression and Melanin Count (in Vivo Experimental Study on Hyperpigmented Wistar Mice Exposed to UVB Exposed Hyperpigmentation)

- 98) Pierrat MJ, Marsaud V, Mauviel A, Javelaud D. Expression of microphthalmia-associated transcription factor (MITF), which is critical for melanoma progression, is inhibited by both transcription factor GLI2 and transforming growth factor- $\beta$ . *Journal of Biological Chemistry*. 2012;287(22):17996–8004.
- 99) Kim SS, Kim MJ, Choi YH, Kim BK, Kim KS, Park KJ, et al. Down-regulation of tyrosinase, TRP-1, TRP-2 and MITF expressions by citrus press-cakes in murine B16 F10 melanoma. *Asian Pac J Trop Biomed*. 2013;3(8):617–22.
- 100) Nishioka E, Funasaka Y, Kondoh H, Chakraborty AK, Mishima Y, Ichihashi M. Expression of tyrosinase, TRP-1 and TRP-2 in ultraviolet-irradiated human melanomas and melanocytes: TRP-2 protects melanoma cells from ultraviolet B induced apoptosis. Vol. 9, *Melanoma Research*. 1999. p. 433–43.



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